

Asymmetric Synthesis of Polyhydroxy Pyrrolidinonyl Nucleoside Analogues from Tartaric acid

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Abstract: Asymmetric synthesis of novel optically active nucleoside analogues **7** from natural tartaric acid is described. In the given nucleoside analogues an optically active polyhydroxy pyrrolidinonyl ring is in place of the tetrahydrofuran ring.

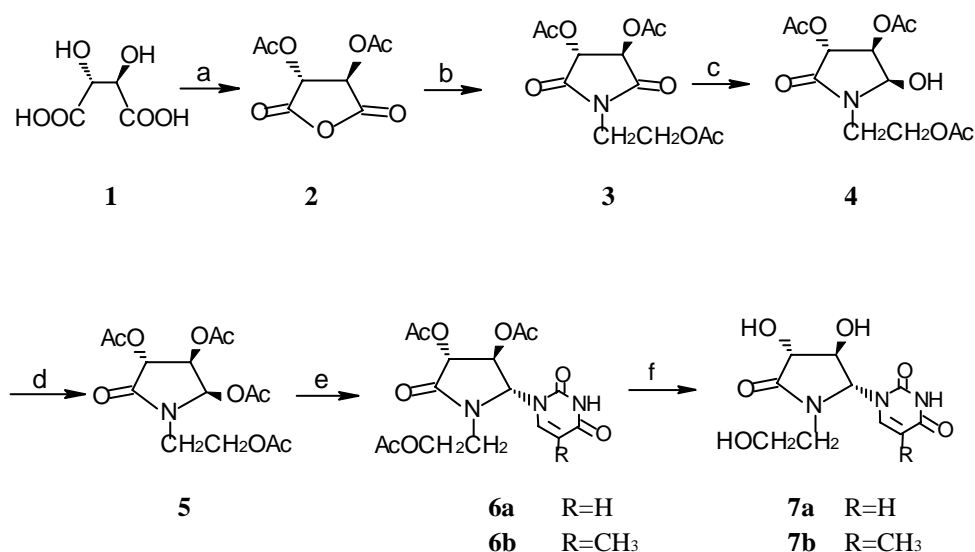
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Modification of nucleoside is an efficient procedure to develop new potent agents against human tumor or viruses¹. More challenging is to synthesize new optically active polyhydroxy nucleoside analogues. Because of the limitation of resources, it seems a rather arduous work to synthesize optically active carbocyclic or other heterocyclic nucleoside analogues with more than two chiral carbons, though natural sugars are available starting materials to oxa-cyclic nucleosides, such as furanosyl or pyranosyl ones. In this paper, we report an efficient and general synthetic route to optically active polyhydroxy aza-nucleosides from natural tartaric acid.

The synthesis of the pyrrolidinonyl nucleoside analogues **7** is shown in **scheme 1**. Reflux of a suspension of L-tartaric acid in acetyl anhydride gave diacetoxy succinic anhydride **2**². The anhydride **2** was treated with 2-aminoethanol (2 eq) in CH₂Cl₂ at room temperature and successively in acetyl chloride at reflux to form (3*R*, 4*R*)-3,4-diacetoxy succinimide **3** in 80% yield³. The excess of free amino group was kept to avoid the possible acylation of the hydroxy group by addition of the anhydride **2** to the solution of aminoethanol in dichloromethane. One mole excess of aminoethanol was used as a base which could be replaced by triethylamine. In the presence of excessive aminoethanol, 2-diacetyl aminoethyl acetate was formed which could be separated from the desired product by chromatography on silica gel. Diastereoselective reduction of **3** with sodium borohydride⁴ in methanol afforded (3*R*, 4*R*, 5*R*)-3,4-diacetoxy-1-(2-acetoxyethyl)-5-hydroxy-2-pyrrolidinone **4**; the result obtained seems different from that given by Yuda⁵. The diastereoselectivity (about 95%) of the reduction was determined based on the ¹H NMR data of **5**, which was derived from **4** via acylation of **4** with acetic anhydride/pyridine in quantitative yield. The configurational assignment of **5** was made by the observed vicinal coupling constants (*J*₃₋₄=4.3Hz,

$J_{4-5}=2.2\text{Hz}$). Additional support on the conclusion of the *cis*-diastereoselective reduction of **3** with sodium borohydride comes from the evidence of the reduction of the compounds from L-malic acid⁶. Condensation⁷ of **5** with *bis*-(trimethylsilyl)uracil or *bis*-(trimethylsilyl)thymine⁸ in the presence of TiCl_4 at -15°C afforded protected pyrrolidinonyl nucleoside analogues **6** in 60% yield. The vicinal coupling constants ($J_{3-4}=4.3\text{Hz}$, $J_{4-5}=6\text{Hz}$) of **6** indicated the *trans*-diastereoselectivity of the condensation. The conclusion of the configurational assignment is in accord with that given by Langlois⁹, although a different result was reported by Yuda⁵. Deacylation¹⁰ of **6** with ammonia in methanol at 5°C gave the final pyrrolidinonyl nucleoside analogues **7** in 90% yield. It was detected that the acetyl group in pyrrolidinonyl ring was removed prior to that in the side chain. Completion of the deacylation was monitored by TLC (eluent: dichloromethane/methanol, 95/5).

Scheme 1. The synthetic route of the compounds **7**



Reagents, conditions and yields:

- (a) acetic anhydride, reflux, 2 hrs, 90%;
- (b) 2-aminoethanol/ CH_2Cl_2 , then CH_3COCl , reflux, 5 hrs, 80%;
- (c) $\text{NaBH}_4/\text{CH}_3\text{OH}$, $-15^\circ\text{C} \sim -5^\circ\text{C}$, 10 mins, 88%;
- (d) $\text{Ac}_2\text{O}/\text{Py}$, 2 hrs, quantitatively;
- (e) *bis*-(trimethylsilyl)uracil or *bis*-(trimethylsilyl)thymine/ $\text{TiCl}_4/\text{CH}_3\text{CN}$, $-20 \sim -10^\circ\text{C}$, 3 hrs, 60%;
- (f) $\text{NH}_3/\text{CH}_3\text{OH}$, 5°C , 3 days, 90%.

Table 1. ^1H and ^{13}C NMR spectral data of the compounds

Compds	^1H , ^{13}C NMR spectral data
3	δ (CDCl_3): 2.0(s, 3H, CH_3), 2.19(s, 6H, 2 CH_3), 3.84(m, 2H, CH_2OCO), 4.21(m, 1H, NCHH), 4.32(m, 1H, NCHH), 5.52(s, 2H, CH in cycle)
4	δ (CDCl_3): 2.06(s, 3H, CH_3), 2.15(s, 3H, CH_3), 2.16(s, 3H, CH_3), 3.59(ddd, $J=4.6 \times 6.6 \times 14.6\text{Hz}$, 1H, H in chain), 3.74(ddd, $J=4.6 \times 6.5 \times 14.6\text{Hz}$, 1H, H in chain), 4.23(ddd, $J=4.6 \times 6.6 \times 11.6\text{Hz}$, 1H, H in chain), 4.3(ddd, $J=4.6 \times 6.5 \times 11.6\text{Hz}$, 1H, H in chain), 5.08(d, $J=2.6\text{Hz}$, 1H, H-3), 5.11(dd, $J=2.6 \times 4.8\text{Hz}$, 1H, H-4), 5.14(d, $J=4.8\text{Hz}$, 1H, H-5)
5	δ (CDCl_3): 2.06(s, 3H, CH_3), 2.12(s, 3H, CH_3), 2.16(s, 3H, CH_3), 2.17(s, 3H, CH_3), 3.26(ddd, $J=4.1 \times 6.8 \times 14.8\text{Hz}$, 1H, H in chain), 3.88(ddd, $J=4.3 \times 6.6 \times 14.8\text{Hz}$, 1H, H in chain), 4.12(ddd, $J=4.3 \times 6.8 \times 11.8\text{Hz}$, 1H, H in chain), 4.36(ddd, $J=4.1 \times 6.6 \times 11.8\text{Hz}$, 1H, H in chain), 5.22(dd, $J=2.2 \times 4.3\text{Hz}$, 1H, H-4), 5.34(d, $J=4.3\text{Hz}$, 1H, H-3), 6.23(d, $J=2.2\text{Hz}$, 1H, H-5)
6a	δ (CDCl_3): 2.07(s, 3H, CH_3), 2.17(s, 3H, CH_3), 2.20(s, 3H, CH_3), 2.96(ddd, $J=2.8 \times 6.3 \times 14.8\text{Hz}$, 1H, H in chain), 3.98(ddd, $J=3.1 \times 6.8 \times 14.8\text{Hz}$, 1H, H in chain), 4.12(ddd, $J=3.1 \times 6.3 \times 11.8\text{Hz}$, 1H, H in chain), 4.32(ddd, $J=2.8 \times 6.8 \times 11.8\text{Hz}$, 1H, H in chain), 5.06(d, $J=4.3$, 1H, H-3), 5.43(dd, $J=4.3 \times 5.9\text{Hz}$, 1H, H-4), 5.9(d, $J=8.0\text{Hz}$, 1H, H in uracil), 6.24(d, $J=5.9\text{Hz}$, 1H, H-5), 7.5(d, $J=8.0\text{Hz}$, 1H, H in uracil), 9.6(bs, 1H, NH).
6b	δ (CDCl_3): 2.05(d, $J=1.2\text{Hz}$, 3H, CH_3), 2.15(s, 3H, CH_3), 2.25(s, 3H, CH_3), 2.29(s, 3H, CH_3), 3.0(ddd, $J=3.1 \times 6.8 \times 15.1\text{Hz}$, 1H, H in chain), 4.08(ddd, $J=3.2 \times 7.0 \times 15.1\text{Hz}$, 1H, H in chain), 4.21(ddd, $J=3.2 \times 6.7 \times 12.1\text{Hz}$, 1H, H in chain), 4.41(ddd, $J=3.1 \times 7.0 \times 12.1\text{Hz}$, 1H, H in chain), 5.15(d, $J=4.3\text{Hz}$, 1H, H-3), 5.53(dd, $J=4.3 \times 6.0\text{Hz}$, 1H, H-4), 6.34(d, $J=6\text{Hz}$, 1H, H-5), 7.35(s, 1H, H in thymine), 8.9(bs, 1H, NH).
7a	δ ($\text{DMSO-d}_6+\text{D}_2\text{O}$): 2.51(m, 1H, H in chain), 3.3~3.44(m, 3H, H in chain), 3.9(bs, 1H, H-4), 4.01(d, $J=6.2\text{Hz}$, 1H, H-3), 5.71(d, $J=8.0\text{Hz}$, 1H, H in uracil), 5.8(bs, 1H, H-5), 7.53(d, $J=8.0\text{Hz}$, 1H, H in uracil).
7b	δ ($\text{DMSO-d}_6+\text{D}_2\text{O}$): 1.80(s, 3H, CH_3), 2.63(m, 1H, H in chain), 3.3~3.5(m, 3H, H in chain), 3.92(bs, 1H, H-4), 4.1(d, $J=6.1\text{Hz}$, 1H, H-3), 5.8(bs, 1H, H-5), 7.3(s, 1H, H in thymine). ^{13}C NMR (DMSO-d_6 , 500MHz) δ : 16.92, 46.79, 62.08, 74.83, 78.88, 81.57, 116.38, 140.06, 156.16, 169.22, 178.86.

The nucleoside analogues synthesized have been characterized using IR, ^1H NMR, at 500MHz and MS(ESI) as well as elemental analysis. The results of biological activity of the compounds prepared will be reported elsewhere.

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